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**United States Patent** [19][11] **Patent Number:** **5,620,850****Bamdad et al.**[45] **Date of Patent:** **Apr. 15, 1997**[54] **MOLECULAR RECOGNITION AT SURFACES DERIVATIZED WITH SELF-ASSEMBLED MONOLAYERS**[75] Inventors: **Cynthia C. Bamdad**, Sharon; **George B. Sigal**, Arlington; **Jack L. Strominger**, Lexington; **George M. Whitesides**, Newton, all of Mass.[73] Assignee: **President and Fellows of Harvard College**, Cambridge, Mass.[21] Appl. No.: **312,388**[22] Filed: **Sep. 26, 1994**[51] **Int. Cl.<sup>6</sup>** ..... **C12Q 1/68**; G01N 33/53; C07K 2/00; C07K 5/00[52] **U.S. Cl.** ..... **530/300**; 435/7.1; 435/7.2; 435/7.9; 435/4; 435/6; 536/388.1; 530/345; 530/400; 422/69; 534/15; 534/16[58] **Field of Search** ..... 435/6, 7.1-7.9, 435/4; 536/388.1; 530/300; 252/700; 422/69[56] **References Cited****U.S. PATENT DOCUMENTS**

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*Primary Examiner*—W. Gary Jones*Assistant Examiner*—Dianne Rees*Attorney, Agent, or Firm*—Wolf, Greenfield & Sacks, P.C.[57] **ABSTRACT**

An article suitable for use as a biosensor includes a species of a formula X—R—Ch adhered to a surface of the article as part of a self-assembled monolayer. X is a functionality that adheres to the surface, R is a spacer moiety, and Ch is a chelating agent. A metal ion can be coordinated by the chelating agent, and a polyamino acid-tagged biological binding partner of a target biological molecule coordinated to the metal ion. A method of the invention involves bringing the article into contact with a medium containing or suspected of containing the target biological molecule and allowing the biological molecule to biologically bind to the binding partner. The article is useful particularly as a surface plasmon resonance chip.

**28 Claims, 4 Drawing Sheets**

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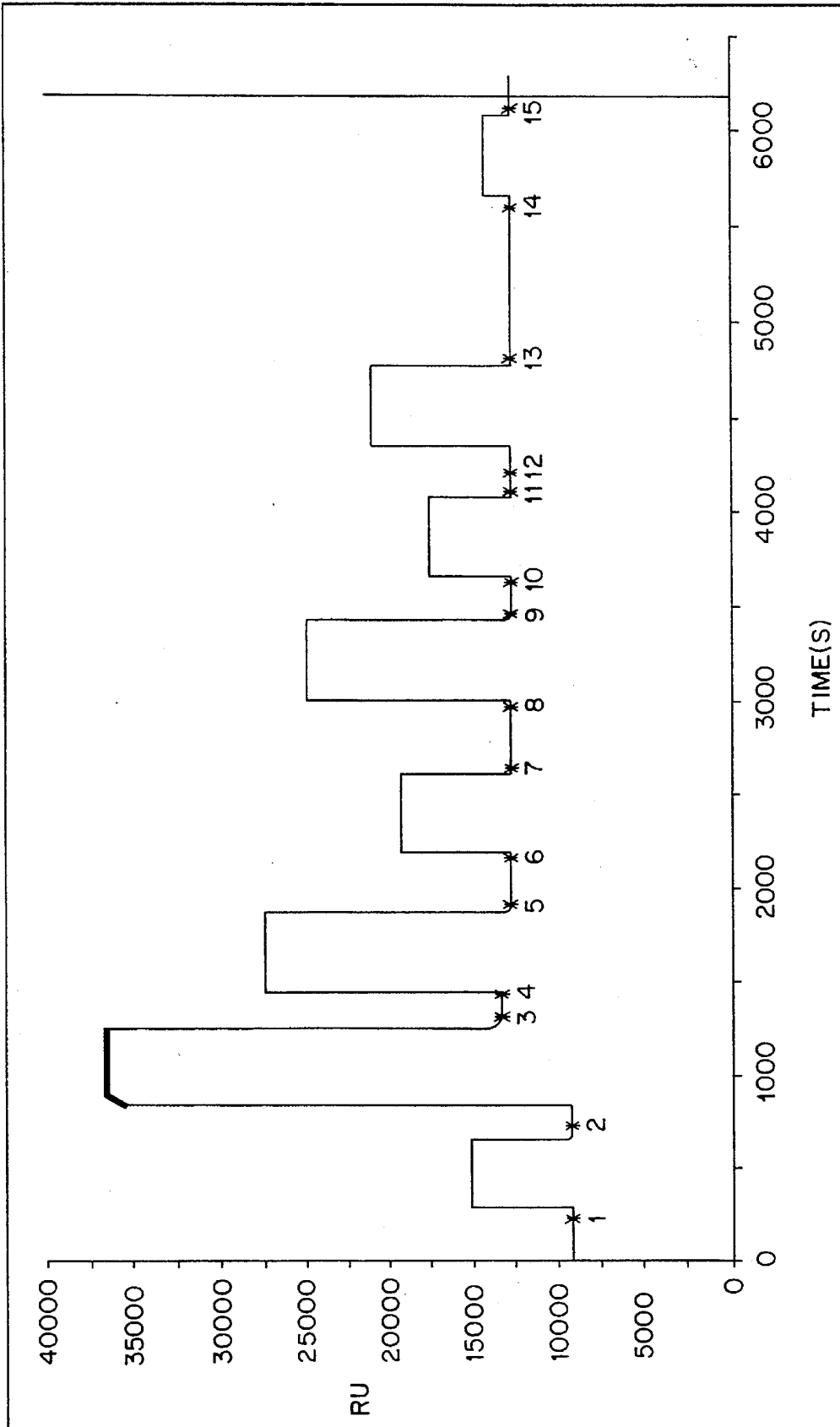


FIG. 1

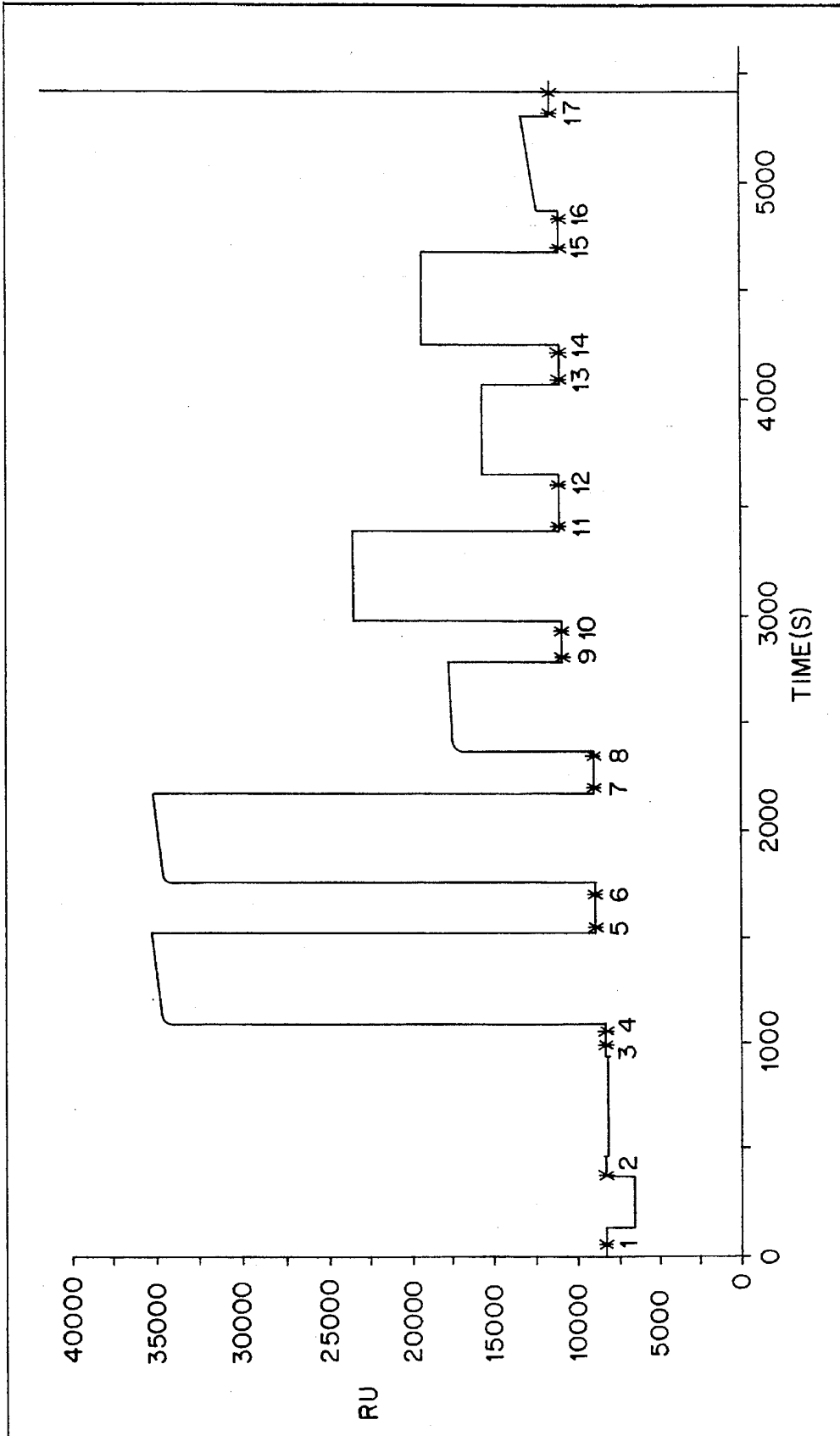


FIG. 2

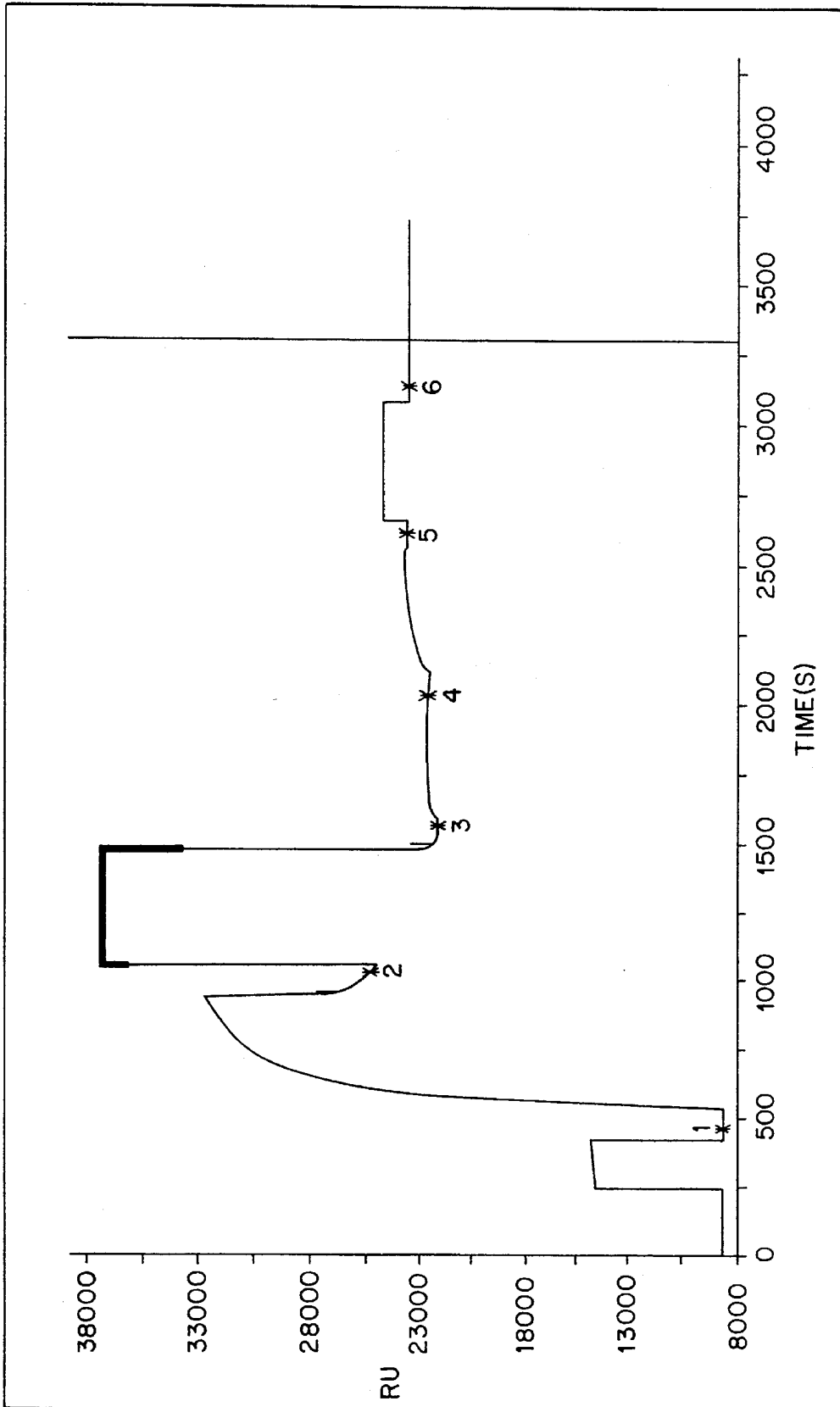


FIG. 3

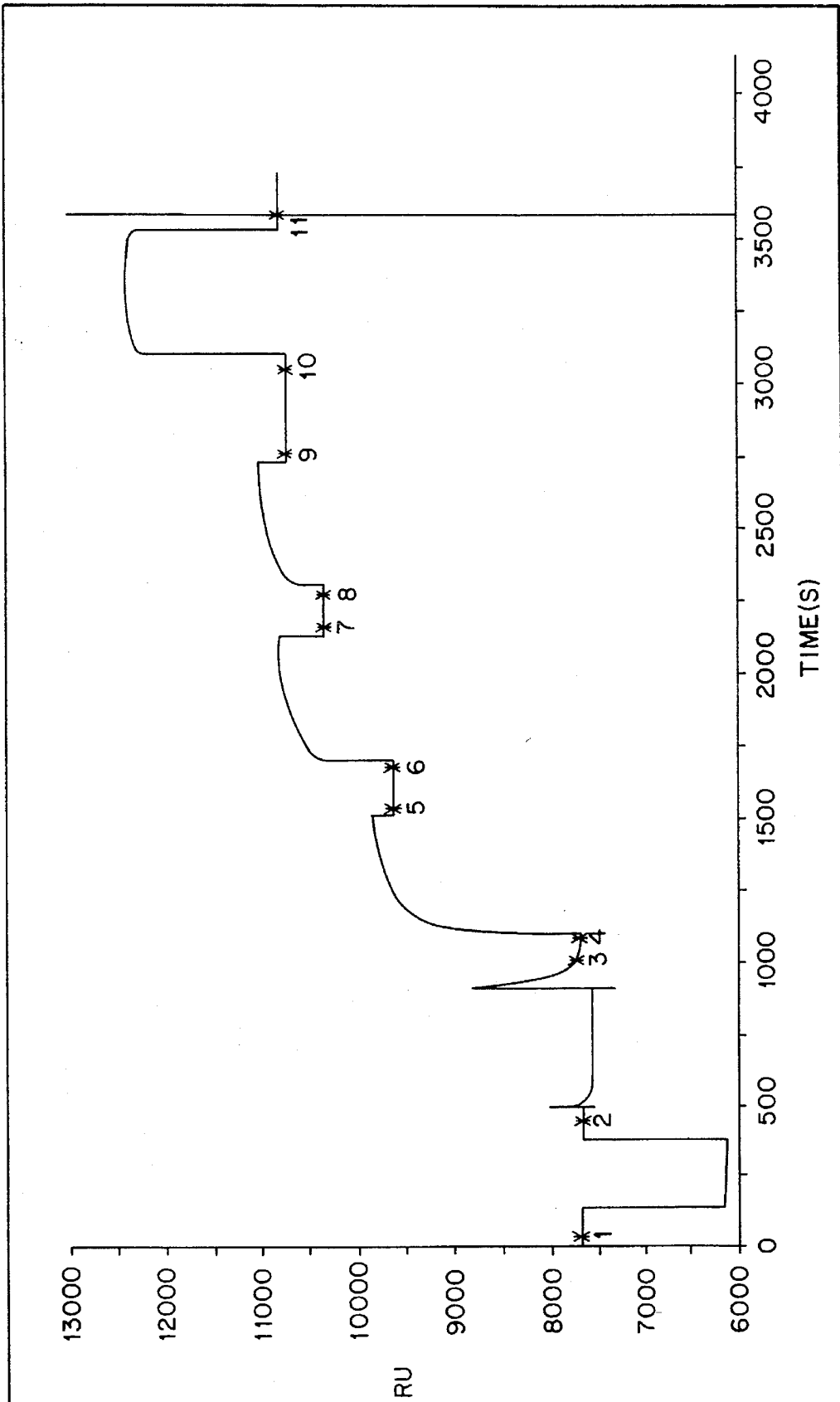


FIG. 4

## MOLECULAR RECOGNITION AT SURFACES DERIVATIZED WITH SELF-ASSEMBLED MONOLAYERS

This invention was supported by NIH grant numbers GM30367 and CA47554. The government has certain rights to the invention.

### FIELD OF THE INVENTION

The present invention relates generally to the derivatization of surfaces for determination of analytes, for example from a fluid medium using a biological binding partner of the analyte. More particularly, the invention relates to the formation on a metal surface of a self-assembled molecular monolayer that exposes a binding partner to an analyte medium in a manner such that analysis of high sensitivity obtains.

### BACKGROUND OF THE INVENTION

Biochemical analyses are invaluable, routine tools in health-related fields such as immunology, pharmacology, gene therapy, and the like. In order to successfully implement therapeutic control of biological processes, it is imperative that an understanding of biological binding between various species is gained. Indeed, an understanding of biological binding between various species is important for many varied fields of science.

Many biochemical analytical methods involve immobilization of a biological binding partner of a biological molecule on a surface, exposure of the surface to a medium suspected of containing the molecule, and determination of the existence or extent of molecule coupling to the surface-immobilized binding partner.

One such technique recently introduced is surface plasmon resonance. This technique utilizes a glass slide having a first side on which is a thin metal film and a second side opposite the first side (known in the art as a sensor chip), a prism, a source of monochromatic and polarized light, a photodetector array, and an analyte channel that directs a medium suspected of containing an analyte to the exposed surface of the metal film. A face of the prism is separated from the second side of the glass slide (the side opposite the metal film) by a thin film of refractive index matching fluid. Light from the light source is directed through the prism, the film of refractive index matching fluid, and the glass slide so as to strike the metal film at an angle at which total internal reflection of the light results, and an evanescent field is therefore caused to extend from the prism into the metal film. This evanescent field can couple to an electromagnetic surface wave (a surface plasmon) at the metal film, causing surface plasmon resonance.

Coupling is achieved at a specific angle of incidence of the light with respect to the metal film (the SPR angle), at which the reflected light intensity goes through a minimum due to the resonance. This angle is determined by a photodetector array as the angle of reflectance and is highly sensitive to changes in the refractive index of a thin layer adjacent to the metal surface. Thus it is highly sensitive to coupling of an analyte to the surface of the metal film. For example, when a protein layer is adsorbed onto the metal surface from an analyte-containing medium delivered to the surface by the analyte channel, the SPR angle shifts to larger values, and this shift is measured by the photodetector array. An article by Stenberg, Persson, Roos, and Urbaniczky entitled, "Quantitative Determination of Surface Concentra-

tion of Protein with Surface Plasmon Resonance Using Radiolabeled Proteins", *Journal of Colloid and Interface Science*, 43: 2, 513-526 (1991), and references therein, describe the technique of surface plasmon resonance. Instrumentation for analysis via surface plasmon resonance is available from Pharmacia Biosensor, Piscataway, N.J., under the trademark BIACore.

Although the introduction of SPR represents an extremely valuable contribution to the scientific community, current state-of-the-art SPR instrumentation lacks the sensitivity needed to detect and analyze certain biological interactions that are at the forefront of scientific inquiry. Experimentation conducted in connection with the instant invention has led to identification of several complications associated with prior art sensor chips, which complications hinder the sensitivity of current SPR techniques. According to one technique for immobilization of a binding partner of an analyte on a surface plasmon resonance sensor chip, long-chain hydroxy-alkyl thiols are adsorbed onto a gold surface as a monolayer, the monolayer's exposed hydroxy groups are activated with epichlorohydrin under basic conditions to form epoxides, a carboxylated dextran gel layer is covalently attached to the monolayer, and a proteinaceous binding partner of an analyte is first electrostatically adsorbed onto the dextran gel layer and then covalently attached thereto. This technique is described in an article by Lofas and Johnsson entitled, "A Novel Matrix on Gold Surfaces in Surface Plasmon Resonance Sensors for Fast and Efficient Covalent Immobilization of Ligands", *J. Chem. Soc., Chem. Comm.* 1526-1528 (1990).

The effectiveness of this approach is hindered by several factors. First, covalent attachment of the proteinaceous binding partner to the gel can affect the binding partner's viability, or activity. Second, covalent attachment of the binding partner to the gel can not be effected with control over the orientation of the binding partner with respect to the surface of the chip (and, importantly, with respect to an analyte-containing medium). Third, non-specific interactions at the gel are promoted by the negative charge that it carries.

According to another technique, a mixed monolayer of hydroxyl and biotin-terminated alkane thiols is prepared on a gold surface, streptavidin is bound to the surface-bound biotin, and biotin-labeled proteins, that are binding partners of analytes, then are attached to empty sites on the streptavidin. However, because biotin must be covalently attached to the protein, this approach lacks control over orientation of the binding partner with respect to the analyte medium, and inactivation of the proteinaceous binding partner due to the formation of covalent linkage can occur. This technique is described in an article by Spinke, Liley, Guder, Angermaier, and Knoll entitled, "Molecular Recognition at Self-Assembled Monolayers: The Construction of Multicomponent Multilayers", *Langmuir*, 9, 1821-1825 (1993).

Accordingly, a general purpose of the present invention is to provide an easily-synthesized chemical species that readily adheres to a surface, and that facilitates surface immobilization of a binding partner of a molecule desirably captured at the surface with a high degree of sensitivity and minimal to zero non-specific binding. It is another purpose of the invention to provide an article with a surface having a high degree of sensitivity for a biological molecule. Another purpose of the invention is to provide a method of capturing a biological molecule, for example at a biosensor surface, by exploiting biological binding interactions that are extremely sensitive to molecular conformation and molecular orientation.

## NOMENCLATURE

The following definitions are provided to facilitate a clear understanding of the present invention.

The term, "chelating agent" refers to an organic molecule having unshared electron pairs available for donation to a metal ion. The metal ion is in this way coordinated by the chelating agent. Two or more neighboring amino acids can act as a chelating agent.

The terms, "bidentate chelating agent", "tridentate chelating agent", and "quadridentate chelating agent" refer to chelating agents having, respectively, two, three, and four electron pairs readily available for simultaneous donation to a metal ion coordinated by the chelating agent.

The term "biological binding" refers to the interaction between a corresponding pair of molecules that exhibit mutual affinity or binding capacity, typically specific or non-specific binding or interaction, including biochemical, physiological, and/or pharmaceutical interactions. Biological binding defines a type of interaction that occurs between pairs of molecules including proteins, nucleic acids, glycoproteins, carbohydrates, hormones and the like. Specific examples include antibody/antigen, antibody/hapten, enzyme/substrate, enzyme/inhibitor, enzyme/cofactor, binding protein/substrate, carrier protein/substrate, lectin/carbohydrate, receptor/hormone, receptor/effector, complementary strands of nucleic acid, protein/nucleic acid repressor/inducer, ligand/cell surface receptor, virus/ligand, etc.

The term "binding partner" refers to a molecule that can undergo biological binding with a particular biological molecule. For example, Protein A is a binding partner of the biological molecule IgG, and vice versa.

The term "biological molecule" refers to a molecule that can undergo biological binding with a particular biological binding partner.

The term "recognition region" refers to an area of a binding partner that recognizes a corresponding biological molecule and that facilitates biological binding with the molecule, and also refers to the corresponding region on the biological molecule. Recognition regions are typified by sequences of amino acids, molecular domains that promote van der Waals interactions, areas of corresponding molecules that interact physically as a molecular "lock and key", and the like.

The term "coordination site" refers to a point on a metal ion that can accept an electron pair donated, for example, by a chelating agent.

The term "free coordination site" refers to a coordination site on a metal ion that is occupied by a water molecule or other species that is weakly donating relative to a polyamino acid tag, such as a histidine tag.

The term "coordination number" refers to the number of coordination sites on a metal ion that are available for accepting an electron pair.

The term "coordinate bond" refers to an interaction between an electron pair donor and a coordination site on a metal ion leading to an attractive force between the electron pair donor and the metal ion.

The term "coordination" refers to an interaction in which one multi-electron pair donor, such as a chelating agent or a polyamino acid tag acting as a chelating agent, coordinatively bonds (is "coordinated") to one metal ion with a degree of stability great enough that an interaction that relies upon such coordination for detection can be determined by a biosensor. The metal ion is coordinated by the multi-electron pair donor.

The term "solid phase" refers to any material insoluble in a medium containing a target molecule or biological molecule that is desirably captured in accordance with the invention. This term can refer to a metal film, optionally provided on a substrate.

The term "surface" refers to the outermost accessible molecular domain of a solid phase.

The term "capturing" refers to the analysis, recovery, detection, or other qualitative or quantitative determination of an analyte in a particular medium. The medium is generally fluid, typically aqueous. The term, "captured", refers to a state of being removed from a medium onto a surface.

The term "target molecule" refers to a molecule, present in a medium, which is the object of attempted capture.

The term "determining" refers to quantitative or qualitative analysis of a species via, for example, spectroscopy, ellipsometry, piezoelectric measurement, immunoassay, and the like.

The term "immobilized", used with respect to a species, refers to a condition in which the species is attached to a surface with an attractive force stronger than attractive forces that are present in the intended environment of use of the surface, and that act on the species. For example, a chelating agent immobilized at a surface, the surface being used to capture a biological molecule from a fluid medium, is attracted to the surface with a force stronger than forces acting on the chelating agent in the fluid medium, for example solvating and turbulent forces.

The term "non-specific binding" (NSB) refers to interaction between any species, present in a medium from which a target or biological molecule is desirably captured, and a binding partner or other species immobilized at a surface, other than desired biological binding between the biological molecule and the binding partner.

The term "self-assembled monolayer" refers to a relatively ordered assembly of molecules spontaneously chemisorbed on a surface, in which the molecules are oriented approximately parallel to each other and roughly perpendicular to the surface. Each of the molecules includes a functional group that adheres to the surface, and a portion that interacts with neighboring molecules in the monolayer to form the relatively ordered array. See Laibinis, P. E.; Hickman, J.; Wrighton, M. S.; Whitesides, G. M. *Science* 245, 845 (1989), Bain, C.; Evall, J.; Whitesides, G. M. *J. Am. Chem. Soc.* 111, 7155-7164 (1989), Bain, C.; Whitesides, G. M. *J. Am. Chem. Soc.* 111, 7164-7175 (1989), each of which is incorporated herein by reference.

The term "self-assembled mixed monolayer" refers to a heterogeneous self-assembled monolayer, that is, one made up of a relatively ordered assembly of at least two different molecules.

## SUMMARY OF THE INVENTION

The foregoing and other objects and advantages of the invention are achieved by providing a molecule having a formula X—R—Ch, in which X represents a functional group that adheres to a gold surface, R represents a spacer moiety that promotes formation of a self-assembled monolayer of a plurality of the molecules, and Ch represents a bidentate, tridentate, or quadridentate chelating agent that coordinates a metal ion. The chelating agent includes a chelating moiety and a non-chelating linker moiety, such that it can be covalently linked via its linker moiety to the



spacer moiety while allowing the chelating moiety to coordinate a metal ion. According to a preferred aspect of the invention a metal ion is coordinated to the chelating agent, and a binding partner of a target molecule is coordinated to the metal ion. This arrangement is facilitated by selecting the chelating agent in conjunction with the metal ion such that the chelating agent coordinates the metal ion without completely filling the ion's coordination sites, allowing the binding partner to coordinate the metal ion via coordination sites not filled by the chelating agent. According to one aspect of the invention the binding partner is a biological species that includes a polyamino acid tag, such as a tag made up of at least two histidine residues, that coordinates the metal ion. In this context the term "adhere" means to chemisorb in the manner in which, for example, alkyl thiols chemisorb to gold.

The present invention also provides a species having a formula  $X-R-Ch-M-BP-BMol$ , in which X represents a functional group that adheres to a surface, R represents self-assembled monolayer-promoting spacer moiety, Ch represents a chelating agent that coordinates a metal ion, M represents a metal ion coordinated by the chelating agent, BP represents a biological binding partner of a biological molecule, and BMol represents the biological molecule. The binding partner is coordinated to the metal ion.

The invention also provides an article including a solid phase that has a surface. A plurality of chelating agents are immobilized at the surface in such a way that essentially each of the chelating agents is oriented so as that the chelating moiety of the agent, that is the electron donating portions of the agent, face in a direction away from the surface and is unencumbered by species, such as other chelating agents, that would interfere with the chelating function. This can be accomplished by isolating the chelating agent at the surface by non-chelating species. In this way each chelating agent can coordinate a metal ion so as to expose in a direction away from the surface at least two free metal coordination sites. According to one aspect of the invention the article includes a surface and a self-assembled mixed monolayer adhered to the surface and formed of at least a first and a second species. The first species has a formula  $X-R-Ch$ , where X, R, and Ch are each selected such that X represents a functional group that adheres to the surface, R represents a spacer moiety that promotes self-assembly of the mixed monolayer, and Ch represents a chelating agent that coordinates a metal ion. The second species is selected to form a mixed self-assembled monolayer with the first species, and the mixed monolayer is made up of at least 70 mol percent of the second species. The second species preferably is a species selected to inhibit non-specific binding of a protein to the surface.

According to a preferred aspect, the article is suitable for capturing a biological molecule. According to this aspect a self-assembled mixed monolayer, formed of a first species and a second species, is adhered to the surface. The first species has a formula  $X-R-Ch-M-BP$ , where X, R, Ch, M, and BP are each selected such that X represents a functional group that adheres to the surface, R represents a spacer moiety that promotes self-assembly of the mixed monolayer, Ch represents a chelating agent that coordinates a metal ion, M represents a metal ion coordinated by the chelating agent, and BP represents a binding partner of the biological molecule. The binding partner is coordinated to the metal ion. The second species is selected to form a mixed, self-assembled monolayer with the first species, and according to a preferred aspect the second species has a formula  $X-R-O-(CH_2CH_2-O)_n-H$ , in which X represents a functional group that

adheres to the surface, R represents a spacer moiety that promotes formation of a self-assembled monolayer of a plurality of the molecules, and n is from one to ten. The article can be constructed and arranged to facilitate instrumental determination of an analyte, and according to a preferred aspect is a biosensor element such as a SPR chip.

The present invention also provides a method of making an article for capturing a target molecule. The method of making the article includes formulating a solution containing a mixture of at least a first and a second species, and exposing to the solution a surface of the article for a period of time sufficient to form a self-assembled mixed monolayer of the first and second species on the surface. The first species has a formula,  $X-R-Ch$  as described above. The second species is selected to form a mixed self-assembled monolayer with the first species, and the second and first species are present in the solution at a molar ratio of at least 70:30.

The present invention also provides a method of capturing a biological molecule. The method involves contacting a medium suspected of containing the biological molecule with a solid phase that has a surface carrying a plurality of binding partners of the biological molecule, in which essentially all of the binding partners are oriented to expose away from the surface a recognition region for the biological molecule. The biological molecule then is allowed to biologically bind to the binding partner, and the biological molecule bound to the binding partner then can be determined. According to one aspect the method involves providing a solid phase having a surface, a chelating agent immobilized at the surface, a metal ion coordinated by the chelating agent, and a biological binding partner of the biological molecule coordinated to the metal ion. According to this aspect the surface is brought into contact with a medium suspected of containing the biological molecule for a period of time sufficient to allow the biological molecule to biologically bind to the binding partner.

The present invention provides another method of capturing a biological molecule. The method involves providing a solid phase having a surface and a metal ion, immobilized at the surface in such a way that the metal ion has at least two free coordination sites. A biological binding partner of a biological molecule is coordinated to the metal ion via a polyamino acid tag, and a medium containing the biological molecule is brought into contact with the surface, whereupon the biological molecule is allowed to biologically bind to the binding partner. The biological molecule then can be determined.

The present invention provides yet another method of capturing a biological molecule. This method involves providing a solid phase that has a surface having adhered thereto a species having a formula  $X-R-Ch-M-BP$ , in which X represents a functional group that adheres to the surface, R represents a self-assembled monolayer-promoting spacer moiety, Ch represents a chelating agent that coordinates a metal ion, M represents a metal ion coordinated by the chelating agent, and BP represents a binding partner of the biological molecule, coordinated to the metal ion. A target molecule then is allowed to biologically bind to the binding partner. The biological molecule then can be determined, for example by detecting a physical change associated with the surface.

An article provided in accordance with the invention can be a biosensor element, such as a SPR chip, and the determination carried out by measuring surface plasmon resonance associated with the chip. The methods of the

invention that involve capturing a molecule can involve removal of a preselected molecule, such as a biological molecule, from a fluid medium.

The present invention also provides sensing elements fashioned as described above and suitable for use in a biosensor, for determination of a biological molecule and in particular a molecule that is a binding partner of a nucleic acid strand. A particularly preferred sensing element includes a substrate, a metal film having a surface, and a self-assembled monolayer of a species  $X-R-NA-NAB$ .  $X$  represents a functional group that adheres to the surface,  $R$  represents a spacer moiety that promotes formation of a self-assembled monolayer of a plurality of the species,  $NA$  represents a nucleic acid strand, and  $NAB$  represents a nucleic acid strand that is a binding partner of  $NA$  and a binding partner of the biological molecule to be determined.

The present invention also provides a kit including an article having a surface and a molecule  $X-R-Ch$ , both as described above. The kit can include  $M$  and  $BP$ , either separately or combined as species  $X-R-Ch-M$  or  $X-R-Ch-M-BP$ , where  $X$ ,  $R$ ,  $Ch$ ,  $M$ , and  $BP$  are as described herein. The kit also can include  $X-R-NA$ , optionally with  $NAB$ , or  $X-R-NA-NAB$  as described herein.

Another aspect of the invention is the article formed when the foregoing molecule(s) is adhered to a gold surface. In this embodiment the article has a chelating agent as described above attached to a spacer moiety as described above which in turn is adhered via  $X$ .

Other advantages, novel features and objects of the invention will become apparent from the following detailed description of the invention when considered in conjunction with the accompanying drawings.

#### BRIEF DESCRIPTION OF THE DRAWINGS

In the drawings:

FIG. 1 is a SPR sensorgram illustrating response of a prior art chip carrying Gal 11 to SRB2;

FIG. 2 is a SPR sensorgram illustrating response of a chip derivatized in accordance with the present invention, and carrying Gal 11, to SRB2;

FIG. 3 is a SPR sensorgram illustrating response of a prior art chip carrying a T cell receptor (ABC) to monoclonal antibodies C1 and BFI; and

FIG. 4 is a SPR sensorgram illustrating response of a chip, derivatized in accordance with the present invention and carrying a T cell receptor (ABC), to monoclonal antibodies C1 and BFI.

#### DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

The present invention provides, according to one aspect, a method of capturing a biological molecule at a surface, using a biological binding partner of the molecule that is immobilized at the surface. In the case of many biological molecules, biological binding is extremely sensitive to orientation and conformation of the members involved in the binding, that is, the biological molecule and the binding partner. Accordingly, the manner of presentation of the binding partner at the surface to a medium containing the biological molecule is directly related to the sensitivity, and success, of the technique.

Accordingly, one method of the present invention provides a chelating agent immobilized at a surface of a solid phase, a metal ion coordinated by the chelating agent, and a biological binding partner of the biological molecule coordinated to the metal ion. The biological binding partner includes a polyamino acid tag that coordinates the metal ion, and a recognition region for the biological molecule remote from the polyamino acid tag. A medium containing the biological molecule is brought into contact with the surface for a period of time sufficient to allow the biological molecule to biologically bind to the binding partner.

The chelating agent and metal ion are selected such that at least two free coordination sites remain on the metal ion after coordination by the chelating agent, and the chelating agent is oriented so as to expose away from the surface the free coordination sites. It is especially preferred that the at least two free coordination sites of the metal are adjacent to each other. Thus, a biological binding partner that includes both a polyamino acid tag that coordinates the metal ion, and a recognition region for the biological molecule that is remote from the polyamino acid tag, is readily immobilized at the surface by coordination to the ion. In this state the binding partner exposes away from the surface the recognition region. In accordance with the method, superior biological binding is observed between a biological molecule in a medium contacted with the surface, and the surface-immobilized binding partner.

The present invention provides an article that can facilitate the above and other methods of the invention. The article includes a solid phase having a surface and a self-assembled mixed monolayer adhered to the surface, the mixed monolayer formed of at least a first species and a second species. The first species has a formula  $X-R-Ch$ , in which  $X$ ,  $R$ , and  $Ch$  are each selected such that  $X$  represents a functional group that adheres to the surface,  $R$  represents a spacer moiety that promotes self-assembly of the mixed monolayer, and  $Ch$  represents a chelating agent that coordinates a metal ion. The second species is selected to form a mixed self-assembled monolayer with the first species, and the molar ratio of the second to the first species is at least 70:30. According to a preferred embodiment, the molar ratio is at least 80:20, according to a more preferred embodiment the molar ratio is at least 90:10, and according to a particularly preferred embodiment, the mixed monolayer is made up of about 95 mol % of the second species and about 5 mol % of the first species. A variety of spectroscopic techniques are available to those of ordinary skill in the art for determining a final ratio of first and second species immobilized at a surface.

According to an aspect of the invention particularly useful in the above-described and other articles and methods, a metal ion is coordinated to the chelating agent, and the metal ion and chelating agent are selected so that at least two coordination sites of the metal remain free after coordination by the chelating agent. One embodiment of this aspect of the invention is facilitated by the above-described molar ratio of the second to the first species of the invention, as discussed below.

A mixed monolayer of the first and second species in the molar ratio of the invention is important to the superior sensitivity of the article. The molar ratio provides for isolation of each chelating agent immobilized at the surface. That is, each chelating agent is separated from other chelating agents by a distance sufficient to assure that each metal ion presented to the surface is coordinated by one chelating agent only, rather than being tethered by coordinate bonds to two or more chelating agents. This is important for several

reasons, including the following. When each metal ion is coordinated by one chelating agent, the number of free coordination sites on the metal available for further coordinate bonding is controlled. That is, selection of a chelating agent and a metal ion can be made in such a manner that the ratio of chelating agents to metal ions at the surface is essentially 1:1, and a desired number of free coordination sites exist on each metal after coordination by the chelating agent. In contrast, if chelating agents are present at the surface in sufficient proximity to each other such that more than one chelating agent can form coordinate bonds with a single metal ion, the metal ion can be coordinated at more than a desired number of coordination sites by the multiple chelating agents, leaving less than a desired number of free coordination site for further coordinate bonding. Additionally, in such a situation control over orientation of the any free coordination sites on the metal ion can be lost; free coordination sites may not face away from the surface, but may face in a direction inaccessible by a species desirably subsequently coordinated to the metal ion at the surface. Further, in such a situation in which a chelating agent can form a coordinate bond with more than one metal ion, one or more electron pairs of a chelating agent available for donation can remain uninvolved in coordinate bonding, which can lead to NSB at the surface. In short, the molar ratio of the second to the first species provides control such that the metal ion is coordinated as stably as possible (lowest possible energy state between chelating agents and metal ions), is oriented with free coordination sites facing away from the surface, and NSB at the surface is minimized or eliminated.

According to one aspect of the invention, the second species is selected to expose to the medium containing the biological molecule a functionality that inhibits NSB. Specifically, a functionality is selected to inhibit NSB of species present at relatively high concentration in a medium in which the biological molecule is presented to the surface. That is, the second species is selected among those that include chemical functionality, at the end of the molecule opposite the functionality that adheres to the surface, that does not bind species in the medium (the end opposite the functionality that adheres to the surface is presented to the medium, or "exposed" at the surface).

The second species has a formula  $X-R-NSBi$ , where NSBi is a NSB-inhibitor. NSBi can be selected from such groups as  $-CH_3$ ;  $-OH$ ;  $-O(CH_2)_nH$ , where  $n=1-15$ , preferably 1-7;  $-CONH(CH_2)_nH$ , where  $n=1-15$ , preferably 1-7;  $-NHCO(CH_2)_nH$ , where  $n=1-15$ , preferably 1-7;  $-(OCH_2CH_2)_nH$ , where  $n=1-15$ , preferably 1-7;  $-COOH$ ;  $-O(CH_2CH_2-O)_nH$  (where  $n=1-10$ );  $-(CF_2)_nCF_3$ , where  $n=1-20$ , preferably 1-8; olefins, and the like. Preferred are  $-CH_3$ ,  $-OH$ , and  $-O(CH_2CH_2-O)_nH$ . According to a preferred embodiment in which a medium containing biological, particularly proteinaceous, species contacts the surface, the second species preferably has a formula  $X-R-O-(CH_2CH_2-O)_n-H$ , in which X represents a functional group that adheres to the surface, R represents a spacer moiety that promotes formation of a self-assembled monolayer of a plurality of the molecules, and n is from one to ten. According to a more preferred embodiment, n is from 2 to 5, and according to a particularly preferred embodiment, n is 3.

The surface, and the functional group X that adheres to the surface, can be selected among a wide variety known to those of skill in the field of surface science. A nonlimiting exemplary list of combinations of surface materials and functional groups X suitable for use in the present invention

follows. Although the following list categorizes certain preferred materials with certain preferred functional groups which firmly bind thereto, many of the following functional groups would be suitable for use with exemplary materials with which they are not categorized, and any and all such combinations are within the scope of the present invention. Preferred surface materials include metals such as gold, silver, copper, cadmium, zinc, palladium, platinum, mercury, lead, iron, chromium, manganese, tungsten, and any alloys of the above with sulfur-containing functional groups X such as thiols, sulfides, disulfides, and the like; doped or undoped silicon with silanes and chlorosilanes; metal oxides such as silica, alumina, quartz, glass, and the like with carboxylic acids; platinum and palladium with nitriles and isonitriles; and copper with hydroxamic acids. Additional suitable functional groups include acid chlorides, anhydrides, sulfonyl groups, phosphoryl groups, hydroxyl groups and amino acid groups. Additional surface materials include germanium, gallium, arsenic, and gallium arsenide. Additionally, epoxy compounds, polysulfone compounds, plastics and other polymers may find use as a surface material in the present invention. Additional materials and functional groups suitable for use in the present invention can be found in U.S. Pat. No. 5,079,600, issued Jan. 7, 1992, incorporated herein by reference.

According to a more preferred embodiment, a combination of gold as surface material and a functional group X having at least one sulfur-containing functional group such as a thiol, sulfide, or disulfide is selected. The interaction between gold and such sulfur-containing functional groups is a well-studied science, and a nonlimiting representative exemplary list of such sulfur-containing functionalities may be found in an article entitled "Wet Chemical Approaches to the Characterization of Organic Surfaces: Self-Assembled Monolayers, Wetting and the Physical-Organic Chemistry of the Solid-Liquid Interface", by G. W. Whitesides and Paul E. Laibinis, *Langmuir*, 6, 87 (1990), incorporated herein by reference. Particularly preferred in the present invention is a gold surface, and a thiol as functional group X.

The spacer moiety R can be selected from among organic spacer moieties that promote formation of self-assembled monolayers. Such moieties are well-known to those of ordinary skill in the art, as described in the above-referenced articles by Laibinis, et al. *Science* 245, 845 (1989), Bain, et al. *J. Am. Chem. Soc.* 111, 7155-7164 (1989), and Bain, C, et al. *J. Am. Chem. Soc.* 111, 7164-7175 (1989). Preferred moieties R are hydrocarbon chains optionally interrupted by hetero groups, of a length of at least eight carbon atoms. As used herein, "hydrocarbon" is meant to define includes alkyl, alkenyl, alkynyl, cycloalkyl, aryl, alkaryl, aralkyl, and the like. Hetero groups can include  $-O-$ ,  $-CONH-$ ,  $-CONHCO-$ ,  $-NH-$ ,  $-CSNH-$ ,  $-CO-$ ,  $-CS-$ ,  $-S-$ ,  $-SO-$ ,  $-(OCH_2CH_2)_nR$  (where  $n=1-20$ , preferably 1-8),  $-(CF_2)_n-$  (where  $n=1-20$ , preferably 1-8), olefins, and the like. It is important that the R is a self-assembled monolayer-promoting moiety. Whether or not a particular moiety promotes formation of a self-assembled monolayer can be routinely determined by one of ordinary skill in the art, optionally in accordance with the teachings of the preceding references, using for example surface spectroscopic techniques.

According to a preferred embodiment,  $R=-(CH_2)_n-$ , where n is from about 8 to about 24, preferably from about 10 to about 20, most preferably from about 9 to about 16. According to an embodiment of the invention in which R is a moiety in the group  $X-R-Ch$ , R preferably comprises  $-(CH_2)_n-O(CH_2CH_2-O)_m-$ , where  $n=4-20$ , preferably

8–14, and  $m=1-10$ , preferably 2–5. A variety of moieties R can be used on different molecules forming a self-assembled mixed monolayer, so long as other requirements described herein are met.

R should also be selected to be chemically stable to reagents used in the synthesis of a species into which it is incorporated. For example, if the species is formed by cleavage of a disulfide, R should be stable to reagents such as mercaptoethanol and dithiothreitol.

the metal ion is preferably selected from those that have at least four coordination sites, preferably six coordination sites. A non-limiting list of metal ions suitable includes  $\text{Co}^{3+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Pd}^{2+}$ ,  $\text{Pt}^{2+}$ ,  $\text{Pd}^{4+}$ ,  $\text{Pt}^{4+}$ ,  $\text{Rh}^{3+}$ ,  $\text{Ir}^{3+}$ ,  $\text{Ru}^{3+}$ ,  $\text{Co}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Au}^{3+}$ ,  $\text{Au}^{+}$ ,  $\text{Ag}^{+}$ ,  $\text{Cu}^{+}$ ,  $\text{MO}_2^{2+}$ ,  $\text{Tl}^{3+}$ ,  $\text{Tl}^{30}$ ,  $\text{Bi}^{3+}$ ,  $\text{CH}_3\text{Hg}^{+}$ ,  $\text{Al}^{3+}$ ,  $\text{Ga}^{3+}$ ,  $\text{Ce}^{3+}$ ,  $\text{UO}_2^{2+}$ , and  $\text{La}^{3+}$ .

The chelating agent is preferably selected from bidentate, tridentate, and quadridentate, chelating agents, and is selected in conjunction with the metal ion so that when the chelating agent coordinates the metal ion, at least two free coordination sites of the metal remain. The chelating agent and metal are selected so that the chelating agent can coordinate the metal ion with a degree of stability great enough that the metal ion will remain immobilized at the surface by the chelating agent.

Additionally, the chelating agent is selected as one that has a chelating moiety and a non-chelating linker moiety, such that it can be covalently linked via its linker moiety to the spacer moiety R while leaving the chelating moiety undisturbed by the covalent linkage and free to coordinate a metal ion. Alternatively, the chelating agent can be selected as one that can be modified via routine organic synthesis to include a non-chelating linker moiety, if such synthesis leaves undisturbed the chelating moiety. One of ordinary skill in the art will appreciate that the non-chelating linker moiety should provide functionality suitable for chemical linkage such as, for example, an amine, alcohol, carbamate, carboxylic acid, thiol, aldehyde, olefin, etc., for formation of an ester linkage, formation of an amide linkage, thiol displacement and thio ether formation, and the like.

With the above considerations in mind, suitable chelating agents and corresponding metal ions can be selected by those of ordinary skill in the art. In accordance with such selection reference can be made to "Chelating Agents and Metal Chelates", Dwyer, F. P.; Mellor, D. P., Academic Press, and "Critical Stability Constants", Martell, A. E.; Smith, R. M., Plenum Press, New York. These works describe a variety of chelating agents, and discuss the stability of coordination between chelating agents and metal ions. Preferably, a chelating agent and metal ion is selected such that the dissociation constant of the combination in aqueous solution is better than 10 nM at physiological pH, that is, such that at least one half of the metal ions are coordinated by chelating agent at a concentration of 10 nM.

A non-limiting exemplary list of suitable chelating agents includes nitrilotriacetic acid, 2,2'-bis(salicylideneamino)-6,6'-demethyldiphenyl, and 1,8-bis(a-pyridyl)-3,6-dithiaoctane.

In some cases it may be advantageous to test a particular chelating agent/metal ion pair to determine whether coordination will be sufficiently stable for use in the present invention. It is within the realm of routine experimentation to one of ordinary skill in the art to follow the teachings herein to immobilize a chelating agent at a surface, such as at a gold SPR chip surface, and then to test the interaction between the chelating agent and the metal ion via, for

example SPR spectroscopy under various conditions. In addition, preliminary screening can be carried out by reacting a prospective chelating agent and metal ion in solution and analyzing the solution spectroscopically. In this regard, reference can be made to, "Spectroscopy and Structure of Metal Chelate Compounds", Nakamoto, K.; McCarthy, S. J., Wiley, New York.

According to one aspect of the invention, an article suitable for capturing a biological molecule is provided. The article includes a self-assembled mixed monolayer of a first species and the second species as described above. The first species has a formula  $\text{X}-\text{R}-\text{Ch}-\text{M}-\text{BP}$ , where X, R, Ch, and M are as described above, and BP is a binding partner of a biological molecule, coordinated to the metal ion.

According to a preferred embodiment the binding partner includes a recognition region for the biological molecule, and a polyamino acid tag that can coordinate the metal ion and that is remote from the recognition region. A polyamino acid tag is meant to define a series of amino acids in proximity such that they can coordinate the at least two free coordination sites of the metal ion. According to a one embodiment, from 2 to about 10 neighboring amino acids such as, for example, neighboring histidines, lysines, arganines, glutamines, or any combination of these can serve as a polyamino acid tag. According to a preferred embodiment, the polyamino acid tag includes at least two, and more preferably from two to 10 neighboring histidines, and according to a particularly preferred embodiment the polyamino acid tag includes from about 3 to about 8 neighboring histidines. With reference to the above-identified work entitled, "Critical Stability Constants" (Martell, et al), selection of these and additional amino acids can be made in conjunction with selection of a metal ion M.

A variety of vectors that express proteins or fragments thereof containing a histidine tag are commercially available from, for example, Novagen, of Madison, Wis. However, these vectors are designed to code proteins or fragments for metal chelate affinity chromatography. For purposes of metal chelate affinity chromatography, it is not important where on the vector the histidine tag lies. The purpose of the histidine tag in that application is solely to allow the chromatography solid phase to adsorb the protein. Therefore, it is not a priority in the commercialization of histidine-tagged proteins that the tag be placed at a location remote from the recognition region.

Where the binding partner is a polyamino acid, a polyamino acid tag can be expressed at a desired location (remote from the recognition region) in a number of ways known to those of ordinary skill in the art, for example by employing the polymerase chain reaction (PCR) to incorporate a nucleic acid sequence that encodes the polyamino acid tag at the appropriate position. Placement of a polyamino acid tag at a desired location is discussed more fully below.

Modification of a protein or fragment thereof by applying to the protein or fragment a polyamino acid tag at a location remote from the recognition region of the protein or fragment can be accomplished readily by one of ordinary skill in the art using, for example, recombinant technology. According to one method for such modification, a desired protein is grown from DNA that codes for the protein, and an expression vector. The protein is isolated and truncated at various amino acid positions, and the protein's specific active sequence (recognition region) elucidated by randomly mutating the resultant sequences. Alternatively, via sequence homology, a variety of similar proteins that rec-

ognize slightly different species are identified and their amino acid sequences determined. The various sequences are compared using a computer, and regions that are variable between the various proteins identified as recognition regions.

Subsequently a strand of DNA for a the desired protein fragment that is large enough to correctly fold can be sequenced with a polyamino acid tag at a location remote from the recognition region. DNA sequencing is routine in the art via, for example, PCR. With an expression vector, the desired polyamino acid-tagged protein fragment then can be readily grown and isolated.

The desired fragment will coordinate to the metal ion via the polyamino acid tag and the recognition region, remote from the tag, will not face the surface, but will be exposed to the medium containing or suspected of containing the binding partner of the protein fragment. As used herein, the term "remote" is meant to define a situation in which the polyamino acid tag is separated from the recognition region by a distance of at least about 20 amino acids, preferably at least about 40 amino acids.

The polyamino acid tagging technique of the invention can be applied to a variety of polyamino acids such as proteins, antibodies, antigens, polymers, and ligands.

Where the binding partner is a not a polyamino acid, it can be coupled chemically, for example covalently coupled, to a polyamino acid including a polyamino acid tag. In this case the coupling of the polyamino acid is effected on the binding partner at a location remote from the recognition region, and/or the polyamino acid that is coupled to the binding partner has a tag at a location remote from the location of coupling to the binding partner. Synthesis of such a species would be routine to those of ordinary skill in the art.

The present invention also provides a method of making an article having a surface for capturing a target molecule. The method involves formulating a solution containing a mixture of at least the first and second self-assembled monolayer-forming species above, and exposing to the surface the solution for a period of time sufficient to form a self-assembled monolayer on the surface.

The amount of time required to allow the monolayer to spontaneously chemisorb to the surface will vary depending upon the solvent, the surface, and the monolayer-forming species. Typically, the time required will be on the order of hours, and often a 24-hour exposure is carried out to make certain that maximal coverage is obtained. The degree of formation of a monolayer can be determined by routine spectroscopic methods, as well as physical method in some cases, such as contact angle measurements and the like.

Other methods for forming the monolayer on the surface are included, for example those disclosed in copending application Ser. No. 08/131,841, filed Oct. 4, 1994 and entitled, "Formation of Microstamped Patterns on Surfaces and Derivative Articles", incorporated herein by reference.

The present invention also provides a sensing element suitable for use in a biosensor, and especially for use in determining a binding partner of a nucleic acid strand.

It has become evident that substances that interact when one or more are bound to DNA do so in a manner that can be highly dependent on the presence of the DNA. Therefore, detailed studies of intricate and sometimes delicate interactions require the presence of the cognate DNA site(s). One example is the study of the interactions which occur in the regulation of transcription. This work is critical for the understanding of many diseases, such as cancer and AIDS. Another example is in the study the interactions of glyco-

proteins, other saccharides, nucleic acid materials, and potential drugs with their intended DNA target(s).

Accordingly, the sensing element includes a substrate, a metal film on the substrate having a surface, and a self-assembled monolayer of a species  $X-R-NA-NAB$  adhered to the surface. X represents a functional group that adheres to the surface, R represents a spacer moiety that promotes formation of a self-assembled monolayer of a plurality of the species, NA represents a nucleic acid strand, and NAB represents a nucleic acid strand that is a binding partner of NA and a binding partner of the biological molecule to be determined.

Attachment of a wide variety of nucleic acid strands NA to a moiety R in a way that the strand can biologically bind to its nucleic acid binding partner NAB can be accomplished with reference to the teaching of example 11, below. It is to be understood that the procedure given for the preparation of the DNA chip may be applied to the preparation of an analogous RNA chip.

The articles of the present invention can be used for a variety of applications, including biosensing applications, test assays, and the like. The term "test assay" generally refers to any procedure in which a member of a biological binding partner pair is to be captured from a medium in which it is dispersed. For example, "test assay" may be used to describe a diagnostic procedure, analytical procedure, microanalytical procedure, forensic analysis, pharmacokinetic study, cell sorting procedure, affinity chromatogram, industrial or laboratory recovery or analysis of one or more species such as toxins, catalysts, or starting materials or products, and the like. A typical test assay is an immunoassay. Biosensing applications include those such as drug screening, environmental monitoring, medical diagnostics, quality control in the pharmaceutical and food industries, and other areas in which it is advantageous to sensitively determine biological binding between partners.

Additionally, in accordance with an embodiment of the invention in which an article has a surface with a monolayer of  $X-R-Ch-M$  adhered thereto, M can serve as a binding partner to capture species that adhere to M. An exemplary list of such species is disclosed in U.S. Pat. No. 5,250,188, issued Oct. 5, 1993 to Bruening, et al. and incorporated herein by reference.

In all of the embodiments of the invention in which an article is provided, a substrate also may be included in the article. The substrate typically will support a film of material that defines the surface, on a side of the film opposite the side at which a self-assembled monolayer is formed.

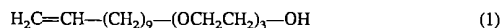
The function and advantage of these and other embodiments of the present invention will be more fully understood from the examples below. The following examples are intended to illustrate the benefits of the present invention, but do not exemplify the full scope of the invention. References cited in the following examples are incorporated herein by reference for all purposes.

#### EXAMPLE 1

##### Preparation of Nitrilotriacetic Acid Chelate Linked Via Spacer Moiety to Thiol

Undec-1-en-11-yl tri(ethyleneglycol) (1) was synthesized according to a procedure reported by Pale-Grosdemange, C.; Simon, E. S.; Prime, K. L.; Whitesides, G. M. *Journal of the American Chemical Society*, 113, 12, (1991).

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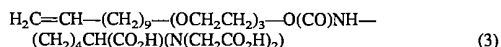


N-(5-amino-1-carboxypentyl)iminodiacetic acid (2) was synthesized according to a procedure reported by Hochuli, E.; Döbeli, H.; Schacher, A. *Journal of Chromatography*, 411, 177 (1987).

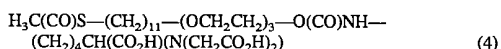


Carbonyldiimidazole (2.3 g, 2 eq. was added to a stirring solution of 2.2 g of alcohol (1) dissolved in 25 ml methylene chloride. After stirring for 2 hours, the solution was applied to a 300 g column of silica equilibrated with ethyl acetate and the imidazole carbamate eluted with 1 liter of ethyl acetate. Evaporation of the solvent under reduced pressure gave 2.7 g (95%) of the imidazole carbamate as an oil.

Amine (2) (5.0 g) was dissolved in 100 ml of water. The pH was titrated to 10.2 with 12N NaOH, then 130 ml of dimethylformamide was added. The imidazole carbamate (2.5 g in 10 ml dimethylformamide) then was added dropwise to aqueous solution of (2) while stirring. After 12 hours, the solution was added to 500 ml water and washed three times with ethyl acetate by gently stirring to avoid the formation of an emulsion. The aqueous phase then was acidified with 6N HCl to pH 1.5 and extracted into ethyl acetate (4×250 ml). The combined extracts were washed with saturated NaCl, dried over MgSO<sub>4</sub>, and the solvent removed under reduced pressure to give 1.8 g (50%) of olefin (3) as a hygroscopic white solid.



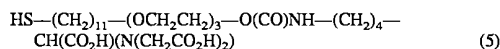
To olefin (3) (1.7 g) dissolved in 15 ml distilled tetrahydrofuran was added 0.5 ml thioacetic acid and 100 mg 2,2'-azobis(2-methylpropionitrile) (AIBN). The solution was irradiated for four hours under a 450 W medium pressure mercury lamp (Ace Glass). The solvent was removed under reduced pressure and the crude product triturated with hexane. Recrystallization from ethyl acetate/hexane gave 1.8 g (94%) of thioacetate as a hygroscopic tan solid (4).



To thioacetate (4) (0.67 g) in 20 ml dimethoxyethane was added 17 ml water, then 20 mg I<sub>2</sub>. After the addition of 3 ml of 2N NaOH, the solution was stirred for four hours while bubbling with O<sub>2</sub>. Addition of 100 ml of water and acidification to pH 1.5 with 6N HCl lead to precipitation of the product as the disulfide. The disulfide was filtered, washed with water and dried under vacuum to give 0.55 g (89%) of a white powder.

The disulfide was reduced to the thiol (6) with triethylphosphine. To the disulfide (0.42 g) in 18 ml methanol plus 2 ml water under an atmosphere of nitrogen, was added 0.3 g of triethylphosphine. The solution was stirred for five hours then concentrated to an oil under reduced pressure. The residue was dissolved in 30 ml degassed water and acidified to pH 1.5 with 6N HCl. The product was extracted three times with 20 ml ethyl acetate. The combined organic phases were washed with saturated NaCl, dried over MgSO<sub>4</sub>, and concentrated under reduced pressure to thiol-linked nitrilotriacetic acid chelate (5) as a tan hygroscopic solid (0.24 g, 57%).

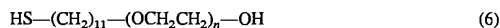
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## EXAMPLE 2

## Preparation of SPR Chip Derivatized With Self-Assembled Mixed Monolayer of Chelate and Nonspecific Binding Inhibitor

11-mercaptoundec-1-yl oligo(ethyleneglycol) (6) was synthesized according to a procedure reported by Pale-Grosdemange, et al., *JACS* (1991; above).

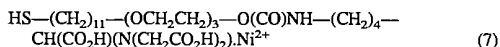


The gold surface of an SPR chip from Pharmacia Biosensor, Piscataway, N.J. was cleaned using 70:30 H<sub>2</sub>SO<sub>4</sub>/H<sub>2</sub>O<sub>2</sub> to remove all species on the gold. 11-mercaptoundec-1-yl oligo(ethyleneglycol) (6) and the thiol-linked nitrilotriacetic acid chelate (5) described in example 1 were dissolved in ethanol in a 95:5 molar ratio, at a total concentration of 1 mM. The chip surface was exposed to the 1 mM solution of the thiols in ethanol for 24 hours, then washed with 95% ethanol and allowed to dry. The derivatized surface was characterized by X-ray photoelectron spectroscopy, which revealed that a mixed monolayer had adhered to the surface, the mixed monolayer including a ratio of (6) and (5) approximately equal to the concentration of those species in the solution from which the surface was derivatized (95:5 molar ratio of (6) to (5)).

## EXAMPLE 3

## Incorporation of Metal Cation Into Chelate of Self-Assembled Monolayer on SPR Chip

The surface derivatized as described in example 2 to have adhered a self-assembled mixed monolayer formed of a 95:5 molar ratio of species (6) and (5) was further modified to allow the chelate to coordinate nickel dication (Ni<sup>2+</sup>). The mixed monolayer was washed with 1 mM aqueous NaOH followed by 1% aqueous Ni(SO<sub>4</sub>).6H<sub>2</sub>O, resulting in species (5) coordinating Ni<sup>2+</sup> to define species (7).



## EXAMPLE 4

## Coordination of Biological Binding Partner To Metal Cation

Gal 11 has been identified as a protein required for efficient galactose utilization in yeast. Nogi, Y.; Fukasawa, T. *Curr. Genet.* 2, 115-120 (1980). Gal 11p (potentiator) is a 44 kD mutant of Gal 11, differing from Gal 11 at a single amino acid only *Id*. Gal 11 is a suspected biological binding partner of Gal 4 (see discussion in example 5). A "mini" form of Gal 11p was expressed and purified (residues 261-351+799-1081) with a polyamino acid tag at a location remote from the recognition region of the protein fragment. Specifically, a run of six histidines is expressed at the amino terminus of the protein fragment, and the histidine-tagged Gal 11p was isolated out of cell extracts by absorption on a column containing nickel dication coordinated to a nitrilotriacetic acid derivative. Hochuli, et al. *J. Chromatog.* 411, 177 (1987), Gentz, R.; Chen, C.; Rosen, C. A.; *Proc. Natl. Acad. Sci.* 86, 821 (1989).

The polyamino acid-tagged binding partner was attached selectively to the derivatized surface described in example 3 by coordination to the metal cation via the histidine tag. Specifically, a 0.25 mg/ml aqueous solution of Gal 11p was injected into a delivery channel of a BIAcore™ SPR instrument, Pharmacia Biosensor. The result was a surface carrying a plurality of binding partners (Gal 11p) oriented such that a substantial portion of the recognition regions for Gal 4 face away from the chip surface and are readily available for biological binding with the biological molecule Gal 4.

#### EXAMPLE 5

##### SPR Determination of Gal 4 Using Chip Derivatized With Self-Assembled Monolayer of Alkyl Thiol Terminating In Metal Chelate Coordinated By Polyamino Acid-Tagged Gal 11p

This example examines the sensitivity of the SPR technique for the analysis of the interaction between biological binding partners, with use of an SPR chip prepared in accordance with the present invention. Specifically, the interaction between the transcription promoter Gal 4 and Gal 11p (which contains increased area of hydrophobicity relative to Gal 11), was studied, using a BIAcore™ SPR instrument available from Pharmacia Biosensor, Piscataway, N.J.

It has been reported that in cells deleted for Gal 11, activation by Gal 4 is reduced some 5–20 fold, an effect that can not be accounted for by decreased levels of the activator. *Id*, Himmelfarb, H. J.; Pearlberg, J.; Last, D. H.; Ptashne, M. *Cell* 63, 1299–1309 (1990), Suzuki, Y.; Nogi, Y.; Abe, A.; Fukasawa, T. *Molecular and Cell Biology* 8, 4991–4999 (1988), Long, R. M.; Mylin, L. M.; Hopper, J. E. *Molecular and Cell Biology* 11, 2311–2314 (1991). It is suspected that proteins containing the Gal 4 region that recognizes Gal 11p in vivo interact in vitro with Gal 11p but not with Gal 11 (wild type).

It is also suspected that Gal 4 promotes transcription by facilitating the necessary proximity of a holoenzyme containing Gal 11 with the TATA box on DNA. It has been reported that Gal 4, truncated to completely abolish its activating domain, is transcriptionally active in the presence of the holoenzyme containing Gal 11p (but not in the presence of the holoenzyme containing Gal 11). Accordingly, it has been postulated that Gal 11p interacts hydrophobically with truncated Gal 4 to facilitate transcription.

SPR was used to study the interaction of Gal 4 with the immobilized his-tagged Gal 11p. It was observed that Gal 11p binds Gal 4, but does not bind a number of other transcription promoters. Histidine-tagged wild-type Gal 11, immobilized on the SPR chip in the same manner, did not bind Gal 4. None of the transcription promoters demonstrate NSB on the mixed monolayer. That is, in the absence of Gal 11p immobilized on the surface, no transcription promoters bound to the surface, and in the presence of immobilized Gal 11p, only Gal 4 was bound.

The interaction between Gal 11p and Gal 4 was not observed by co-immunoprecipitation or on an affinity column, demonstrating the importance of the orientation in the presentation of the protein to the analyte solution, achievable in accordance with the present invention.

#### EXAMPLE 6

##### Comparative Attempted SPR Determination of SRB2 Using Standard Dextran Chip Carrying Gal 11

This example examines the sensitivity of the current state-of-the-art SPR chip, which comprises a layer of dextran on the gold SPR chip surface.

In this example, an attempt was made to study the interaction of Gal 11 (wild-type) and the RNA polymerase B suppressor SRB2, a 28 kD member of the holoenzyme described in example 5 (the multi-protein complex which assembles on the DNA at the TATA box and with which a DNA-bound activator must interact in order to activate transcription). Koleske, A. J.; Young, R. A. *Nature*, 368, 466–469 (1994). As discussed in example 5, Gal 11p interacts with Gal 4, thus it is important to find a target of Gal 11 on the holoenzyme. Detection of this type of interaction is difficult because transcription factors generally exhibit low affinity for their targets, such that positive control of transcription is achieved only through the correct assembly of several factors. The 30 kD protein SRB5 was used as a negative control in this example. SRB5 is shown by genetic experiments not to interact with Gal 11 or Gal 11p *Id*.

An SPR chip having a layer of dextran on gold (#BR1000-14) was purchased from Pharmacia Biosensor, and the SPR instrument described above was employed.

FIG. 1 illustrates a sensorgram plotting resonance units (RU) as a function of time associated with this example. The flow rate of sample across the chip surface was 5 ul/min. The sensorgram of FIG. 1 is labeled with reference numerals that correspond to the steps of the experimental protocol below.

1. (t=240 sec) description of chip type; standard dextran
2. (t=731 sec) end of injection of EDC/NHS which activates the carboxylates on the dextran surface.
3. (t=1326 sec) end of injection of 35 ul wild type Gal 11 at 0.025 mg/ml, in NaOAc buffer at pH 4.5.
4. (t=1440 sec) preinjection baseline
5. (t=1928 sec) end of injection of ethanolamine, which blocks the activated carboxylates that did not covalently link to a protein.
6. (t=2179 sec) preinjection baseline
7. (t=2668 sec) end of injection of SRB2 at 0.25 mg/ml diluted with the running buffer PBS.
8. (t=2991 sec) preinjection baseline
9. (t=3472 sec) end of injection of SRB2 at 0.5 mg/ml
10. (t=3639 sec) preinjection baseline
11. (t=4122 sec) end of injection of SRB5 at 0.25 mg/ml
12. (t=4218 sec) preinjection baseline
13. (t=4814 sec) end of injection of SRB5 at 0.5 mg/ml
14. (t=5610 sec) preinjection baseline
15. (t=6128 sec) end of injection of myc- $\alpha$ , a monoclonal antibody which should bind to Gal 11, at 0.25 mg/ml.

Tabulated below are protein absorption response values (AbsResp; RU units) and response values relative to preceding baseline (RelResp; RU units) corresponding to the protocol steps above.

	Time	AbsResp	RelResp
1.	240.00	9150	
2.	731.00	9313	
3.	1326.50	13444	4131
4.	1440.00	13291	3978
5.	1928.00	12950	3637
6.	2179.00	12872	3559
7.	2668.00	12923	51
8.	2991.00	12880	8
9.	3472.00	12976	96
10.	3639.00	12895	15
11.	4122.00	12911	17
12.	4218.00	12887	-8
13.	4814.00	12941	54

-continued

	Time	AbsResp	RelResp
	14. 5610.00	12852	-35
	15. 6128.00	12940	87

4131RUs of Gal 11 bound to the dextran. The interaction of Gal 11 with SRB2 yielded an absorption of 51 RU's at [SRB2]=0.25 mg/ml; at [SRB2]=0.50 mg/ml, the absorption was 96 RUs. By contrast, at [SRB5]=0.25 mg/ml the absorption was 17 RU's, while at [SRB5]=0.5 mg/ml, the absorption was 54 RUs. Lastly, at [myc- $\alpha$ ]=0.25 mg/ml, the absorption was 87 RU's.

Since the negative and positive controls (SRB5 and myc- $\alpha$ , respectfully) gave essentially the same results, no conclusions regarding binding can be reached. This example demonstrates that the current state-of-the-art dextran chip does not provide the sensitivity necessary for determination of biological binding between Gal 11 and SRB2.

## EXAMPLE 7

SPR Determination of SRB2 Using Chip Derivatized With Self-Assembled Monolayer Alkyl Thiol Terminating In Metal Chelate Coordinated By Polyamino Acid-Tagged Gal 11

As in example 5, this example demonstrates the sensitivity of the SPR technique using a chip derivatized in accordance with the present invention. An SPR chip prepared in accordance with example 2 was mounted in the instrument, modification of the chip to chelate nickel dication was effected in accordance with example 3, the modified chip surface was exposed to histidine-tagged Gal 11, and SPR response to introduction to the chip surface of SRB2 and SRB5 was investigated.

FIG. 2 illustrates a sensorgram plotting resonance units as a function of time associated with this example. The flow rate of sample across the chip surface was 5 ul/min. The sensorgram of FIG. 2 is labeled with reference numerals that correspond to the steps of the experimental protocol below.

1. (t=50 s) chip description; 5% Ni ligand
2. (t=390 s) end of injection of 1 mM NaOH
3. (t=993 s) end of injection of 1% NiSO<sub>4</sub>
4. (t=1055 s) preinjection baseline
5. (t=1555 s) end of injection of Gal 11 at 0.025 mg/ml
6. (t=1703 s) preinjection baseline
7. (t=2222 s) end of injection of a second Gal 11 injection at the same concentration to achieve maximum chip coverage.
8. (t=2343 s) preinjection baseline
9. (t=2821 s) end of injection of SRB2 at 0.25 mg/ml
10. (t=2932 s) preinjection baseline
11. (t=3438 s) end of injection of SRB2 at 0.5 mg/ml
12. (t=3610 s) preinjection baseline
13. (t=4116 s) end of injection of SRB5 at 0.25 mg/ml
14. (t=4238 s) preinjection baseline
15. (t=4728 s) end of injection of SRB5 at 0.5 mg/ml
16. (t=4842 s) preinjection baseline
17. (t=5330 s) end of injection of myc- $\alpha$  at 0.25 mg/ml

Tabulated below are protein absorption response values (AbsResp; RU units) and response values relative to pre-

ceding baseline (RelResp; RU units) corresponding to the protocol steps above.

	Time	AbsResp	RelResp
	1. 50.50	8297	
	2. 390.00	8258	
	3. 993.00	8374	
	4. 1055.00	8368	0
	5. 1555.50	9122	753
	6. 1703.00	8887	519
	7. 2222.00	9197	828
	8. 2343.00	9042	673
	9. 2821.00	10998	1956
	10. 2932.00	10927	1885
	11. 3438.00	11174	2132
	12. 3610.00	11090	2048
	13. 4116.00	11138	47
	14. 4238.00	11100	10
	15. 4728.00	11174	73
	16. 4842.00	11129	28
	17. 5330.00	11578	450

In contrast to the prior art embodiment examined in example 6, substantial binding of SRB2 to immobilized Gal 11 was observed. Importantly, no nonspecific binding of SRB5 to Gal 11 was observed. Specifically, 673 RUs of histidine-tagged Gal 11 bound to the surface exposing chelate coordinating nickel dication. The interaction of Gal 11 with SRB2 yielded an absorption of 1956 RUs at [SRB2]=0.25 mg/ml; at [SRB2]=0.50 mg/ml, the absorption increased by an additional 176 RU's, for a total of 2132 RU's of bound protein. By contrast, at [SRB5]=0.25 mg/ml the absorption was 47 RUs, while at [SRB5]=0.5 mg/ml, the absorption was 73 RUs. Lastly, at [myc- $\alpha$ ]=0.25 mg/ml, the absorption was 450 RU's. These results clearly show that the biological molecule SRB2 bound to its binding partner Gal 11. The negative control SRB5 did not bind, and the positive control myc- $\alpha$  did bind.

As additional control experiments: (1) SRB2 was brought into contact with a chip incorporating a metal ion (prepared in accordance with example 3) that did not have coordinated to it a the binding partner Gal 11, and (2) SRB5 was flowed over the chip after coordination of Gal 11 prior to the introduction of SRB2. No significant binding was observed in either case.

This experiment demonstrates that a biosensor surface prepared in accordance with the present invention, which provides biological binding partners oriented to stably expose recognition regions to an analyte solution, is superior to the prior art surface.

## EXAMPLE 8

Comparative Attempted SPR Determination of Monoclonal Antibodies Using Standard Dextran Chip Carrying T Cell Receptor

As in example 5, this example examines the sensitivity of the current state-of-the-art SPR chip, which comprises a layer of dextran on the gold SPR chip surface.

In this example, an attempt was made to study the interaction of a soluble 42 kD chimeric T cell receptor (TCR), herein referred to as ABC, with 2 monoclonal antibodies of MW=155 kD (C1 and  $\beta$ F1). C1 recognizes a conformational epitope thought to be near the active site of the receptor, while  $\beta$ F1 recognizes a linear epitope of the constant domain. Although the structure of a TCR has not yet been solved, by analogy to IG structure, the C1 epitope should reside near the "top" of the protein and the  $\beta$ F1



epitope near the "bottom", where the bottom is defined as the histidine tag of the protein. Probing the TCR with Cl demonstrated the availability of a defined site on the protein. The ratio of Cl:βFl binding is the ratio of correctly:incorrectly folded protein, since Cl was shown to occlude the binding site of βFl when the TCR is bound to a surface (data not shown).

FIG. 3 illustrates a sensorgram plotting resonance units as a function of time associated with this example. The flow rate of sample across the chip surface was 5 ul/min. The sensorgram of FIG. 3 is labeled with reference numerals that correspond to the steps of the experimental protocol below.

1. (t=470 s) end of injection of EDC/NHS
2. (t=1044 s) end of injection of ABC at 0.125 mg/ml in PBS buffer
3. (t=1579 s) end of injection of ethanolamine
4. (t=2047 s) end of injection of Cl at 0.2 mg/ml in PBS
5. (t=2630 s) end of injection of βFl at 0.2 mg/ml in PBS
6. (t=3152 s) end of injection of myc-α at 0.2 mg/ml in PBS

Tabulated below are protein absorption response values (AbsResp; RU units) and response values relative to preceding baseline (RelResp; RU units) corresponding to the protocol steps above.

	Time	AbsResp	RelResp
1.	470.50	8717	
2.	1044.00	25276	16558
3.	1579.00	22216	13499
4.	2047.00	22675	459
5.	2630.00	23728	1512
6.	3152.00	23658	1442

13,499 RU's of ABC bound to the dextran surface. The interaction with Cl led to an absorption increase of 459 RUs, while that with βFl led to an additional 1053 RU's. Finally, the myc-α antibody did not bind. The ratio of Cl:ABC was 1:29. The ratio of Cl:BI was 1:2.3. Since the molecular weight of Cl is 3.7 times that of ABC, the molar ratio of Cl:ABC is 1:108, which implies that only 0.9% of the ABC molecules were recognized by Cl.

This experiment demonstrates that the current state-of-the-art dextran chip does not provide the sensitivity necessary for determination of biological binding between ABC and Cl.

#### EXAMPLE 9

##### SPR Determination of Monoclonal Antibodies Using Chip Derivatized With Self-Assembled Monolayer of Alkyl Thiol Terminating In Metal Chelate Coordinated By Polyamino Acid-Tagged T Cell Receptor

As in examples 5 and 7, this example demonstrates the sensitivity of the SPR technique using a chip derivatized in accordance with the present invention. An SPR chip prepared in accordance with example 2 was mounted in the SPR instrument described above, modification of the chip to chelate nickel dication was effected in accordance with example 3, the modified chip surface was exposed to ABC (example 8) including a histidine tag at the constant domain, and SPR response to introduction to the chip surface of Cl and βFl was investigated.

FIG. 4 illustrates a sensorgram plotting resonance units as a function of time associated with this example. The flow rate of sample across the chip surface was 5 ul/min. The sensorgram of FIG. 4 is labeled with reference numerals that correspond to the steps of the experimental protocol below.

1. (t=42 s) chip description
2. (t=448 s) end of injection of 1 mM NaOH
3. (t=1017 s) end of injection of 1% NiSO<sub>4</sub>
4. (t=1097 s) preinjection baseline
5. (t=1553 s) end of injection of ABC at 0.125 mg/ml; solution contains 0.02M imidazole
6. (t=1696 s) preinjection baseline
7. (t=2177 s) end of injection of Cl at 0.2 mg/ml; solution contains 0.02M imidazole
8. (t=2288 s) preinjection baseline
9. (t=2781 s) end of injection of βFl at 0.2 mg/ml; solution contains 0.02M imidazole
10. (t=3075 s) preinjection baseline
11. (t=3604 s) end of injection of myc-α at 0.2 mg/ml; solution contains 0.02M imidazole

Tabulated below are protein absorption response values (AbsResp; RU units) and response values relative to preceding baseline (RelResp; RU units) corresponding to the protocol steps above.

	Time	AbsResp	RelResp
1.	42.00	7671	
2.	448.50	7660	
3.	1017.50	7730	
4.	1097.00	7695	
5.	1553.50	9640	1945
6.	1696.00	9634	1939
7.	2177.00	10376	742
8.	2288.00	10353	719
9.	2781.00	10761	408
10.	3075.00	10739	386
11.	3604.00	10830	91

1,945 RU's of ABC bound to the Ni surface. The interaction with Cl led to an absorption increase of 719 RU's, while that with βFl led to an additional 386 RU's. Finally, the myc-α antibody binding led to a 91 RU increase. The ratio of Cl:ABC was 1:2.7. The ratio of Cl:βFl was 1.9:1. Since the molecular weight of Cl is 3.7 times that of ABC, the molar ratio of Cl:ABC is 1:10, which implies that 10% of the ABC molecules were recognized by Cl. In separate in vivo inhibition assays, we consistently found that 10% of the ABC was correctly folded. We therefore conclude that in this example 100% of the correctly folded ABC was oriented on the gold chip with the Cl binding site in the accessible "up" position with respect to the chip surface. This is in contrast to 9% retained activity in example 8.

This example demonstrates that essentially all of the binding partners immobilized at the chip surface in accordance with the present invention were oriented so as to expose in a direction away from the chip surface the recognition region of the binding partner for the biological molecule sought to be captured.

#### EXAMPLE 10

##### Fabrication of Chip Derivatized With Self-Assembled Monolayer of Alkyl Thiol Terminating In Biological Binding Partner of DNA, To Expose DNA For Binding Studies

This prophetic example describes preparation of an alkyl thiol terminating at its exposed end in a nucleic acid

sequence, and a surface derivatized to carry a self-assembled monolayer including such species. Specifically, an SPR chip is primed to receive underivatized dsDNA, provided by the user, which has been modified by the generation of an EcoR I site at one end.

A protected, single-stranded oligo is prepared via standard methods, as described in the Gene Assembler manual, Pharmacia Biosensor. The oligo is at least 25 base pairs long, and is a mixed, nonself-complementary sequence, terminated with a dG at the 3' end. The polymer-supported oligo is dried under vacuum overnight.

The 5' dimethoxytrityl (DMT) protecting group is deprotected at pH 3.0 in water for 10 minutes at room temperature. The reaction is stopped by freezing on dry ice, and the residue is filtered. See Ferentz, A.; Keating, T. A.; Verdine, G. L. *J. Am. Chem. Soc.* 115, 9006-9014 (1993)

From 11-mercaptoundec-1-yl oligo(ethyleneglycol) (6; see example 2) is synthesized the S-DMT-protected derivative. See Zervas, L. L.; Photaki, I. *J. Am. Chem. Soc.* 84, 3887-3891 (1962). The 5' hydroxyl of the oligo is activated with carbonyl diimidazole. See Wachter, I.; Jabloski, J. A.; Ramachandran, K. L. *Nucleic Acids. Res.* 14, 7985-7994 (1986).

The protected alkyl thiol is added to the activated oligo in dioxane water for 30 minutes at room temperature. Excess reagent is washed away with dry dioxane, followed by methanol, and the product is dried in a desiccator overnight. The remaining protecting groups and the solid support are deprotected by treating with 25% aqueous ammonia for 6 hours at 60° C. The aqueous ammonia is chilled at 0° for 30 minutes, the polymer support is filtered, and the ammonia evaporated.

The derivatized oligo is purified via 8% PAGE-7M urea, then via G-10 Sephadex chromatography. The oligo is reduced with silver nitrate/DTT and rechromatographed. See Connolly, B. A.; Rider, P. *Nucleic Acids Res.* 12, 4485-4502 (1985).

An oligo of sequence complementary to that made in step 1, with an additional 4 bp's at the 5' end (3'-TTAA-5') is purchased. The purified, derivatized oligo from the previous step is hybridized with the complementary oligo by denaturation at 75° C., and annealed at 55° C. This yields an alkyl thiol attached to a dsDNA, which is terminated with an EcoR I restriction enzyme site. EcoR I is widely available commercially, for example from New England Biolabs.

This DNA ligand is mixed with a self-assembled monolayer-forming, nonspecific binding-inhibiting species, specifically, 11-mercaptoundec-1-yl oligo(ethyleneglycol) (6; see example 2) in a 5:95 molar ratio, respectively. The chip surface is prepared as described in example 2. This completes the preparation of a commercial chip.

#### EXAMPLE 11

##### Modification of Chip Derivatized With Self-Assembled Monolayer of Alkyl Thiol Terminating In Biological Binding Partner of DNA, To Expose DNA For Binding Studies

In this prophetic example, selected dsDNA that the user wishes to present on the chip surface is cut, via standard methods, with EcoR I. The cut DNA is ligated to the DNA already on the chip using DNA ligase (New England Biolabs). The chip is now ready for use in a biosensor.

Advantages of the chip prepared in accordance with examples 10 and 11 follow. The current state-of-the-art chip, available from Pharmacia Biosensor for use in the BIA-core™ SPR instrument, presents a dextran surface to which strepavidin is attached. The user must either derivative the desired DNA with biotin, or purchase biotinylated DNA. In the former case, the reaction is difficult and biotinylation may occur at any number of sites, leading to a lack of control over orientation on the chip. Furthermore, biotinylation may block an interaction site on the DNA. Commercially available biotinylated DNA is very expensive and cannot be obtained in lengths much greater than 70 base pairs. The derivatized DNA then is attached to the chip surface via interaction between the biotin and the strepavidin.

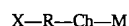
Another disadvantage of the prior art strepavidin/biotin technique includes the fact that many of the compounds which bind negatively charged DNA are positively charged. Since the dextran surface retains a number of negatively charged carboxylates, interactions of an electrostatic nature may be mistaken for binding with the target DNA. Also, kinetic analyses of binding would be compromised due to the additional attractive force created by the negative carboxylates. Still another disadvantage of the prior art technique is that the strepavidin on the chip surface gives rise to nonspecific interactions with many target compounds.

The procedure described in this example for the preparation of the DNA chip may be applied to the preparation of an analogous RNA chip.

Those skilled in the art would readily appreciate that all parameters listed herein are meant to be exemplary and actual parameters will depend upon the specific application for which the methods and apparatus of the present invention are being used. It is, therefore, to be understood that the foregoing embodiments are presented by way of example only and that, within the scope of the appended claims and equivalents thereto, the invention may be practiced otherwise than as specifically described.

What is claimed is:

1. A species having a formula:



wherein X represents a functional group and R represents a spacer moiety that, together, are able to promote formation from solution of a self-assembled monolayer on a surface, Ch represents a chelating agent that coordinates a metal ion, and M represents a metal ion coordinated to the chelating agent, wherein M also is coordinated to a biological binding partner of a biological molecule via coordination sites not filled by the chelating agent or includes coordination sites not filled by the chelating agent that, upon exposure to a polyamino acid tag, are able to become coordinated by the polyamino acid tag.

2. The species as recited in claim 1, wherein the chelating agent is a quadradentate chelating agent.

3. The species as recited in claim 1, wherein the chelating agent is nitrilotriacetic acid.

4. The species as recited in claim 1, wherein the chelating agent and metal ion are selected such that the chelating agent coordinates all but at least two of the metal coordination sites.

5. The species as recited in claim 1, wherein the chelating agent is a tetradentate chelating agent, and the metal ion has a coordination number of six.

6. The species as recited in claim 1, wherein the metal ion is Ni<sup>2+</sup>.

7. The species as recited in claim 1, further comprising a biological binding partner of a biological molecule coordinated to the metal ion.

8. The species as recited in claim 7, wherein the biological binding partner includes a polyamino acid tag that coordinates the metal ion.

9. The species as recited in claim 8, wherein the polyamino acid tag comprises at least two neighboring amino acids defining a chelating agent that coordinates the metal ion.

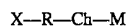
10. The species as recited in claim 9, wherein the at least two neighboring amino acids are at least two neighboring histidines.

11. The species as recited in claim 7, wherein the biological binding partner specifically binds a protein.

12. A species as recited in claim 1, wherein X represents a functional group that adheres to a gold surface.

13. A species as recited in claim 12, wherein X—R—Ch represents a thiol-linked nitrilotriacetic acid chelate.

14. A species having a formula:



wherein X represents a functional group and R represents a spacer moiety that, together, are able to promote formation from solution of a self-assembled monolayer on a gold surface, Ch represents a chelating agent that coordinates a metal ion, selected from the group consisting of bidentate, tridentate, and quadridentate chelating agents, and M represents a metal ion coordinated to the chelating agent, wherein M also is coordinated to a biological binding partner of a biological molecule via coordination sites not filled by the chelating agent or includes coordination sites not filled by the chelating agent that, upon exposure to a polyamino acid tag, are able to become coordinated by the polyamino acid tag.

15. The species as recited in claim 14, wherein the chelating agent is a quadridentate chelating agent, and the metal ion has a coordination number of six.

16. The species as recited in claim 14, wherein the metal ion is Ni<sup>2+</sup>.

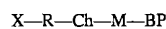
17. The species as recited in claim 14, wherein the chelating agent is nitrilotriacetic acid.

18. A species as recited in claim 14, wherein X—R—Ch represents a thiol-linked nitrilotriacetic acid chelate.

19. The species as recited in claim 14, further comprising a biological binding partner of a biological molecule coordinated to the metal ion.

20. The species as recited in claim 19, wherein the biological binding partner specifically binds a protein.

21. A species having a formula:



wherein X represents a functional group and R represents a spacer moiety that, together, are able to promote formation from solution of a self-assembled monolayer on a surface, Ch represents a chelating agent that coordinates a metal ion, M represents a metal ion coordinated to the chelating agent, and BP represents a biological binding partner of a biological molecule coordinated to the metal ion.

22. The species as recited in claim 21, wherein the biological binding partner includes a polyamino acid tag that coordinates the metal ion.

23. The species as recited in claim 22, wherein the polyamino acid tag comprises at least two neighboring amino acids defining a chelating agent that coordinates the metal ion.

24. The species as recited in claim 23, wherein the at least two neighboring amino acids are at least two neighboring histidines.

25. The species as recited in claim 21, wherein the biological binding partner specifically binds a protein.

26. The species as recited in claim 21, wherein the biological binding partner specifically binds a protein and includes a polyamino acid tag comprising at least two neighboring histidines that coordinates the metal ion.

27. The species as recited in claim 21, wherein X represents a functional group that adheres to a gold surface.

28. A species as recited in claim 21, wherein X—R—Ch represents a thiol-linked nitrilotriacetic acid chelate.

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